

# BioCode® GPP

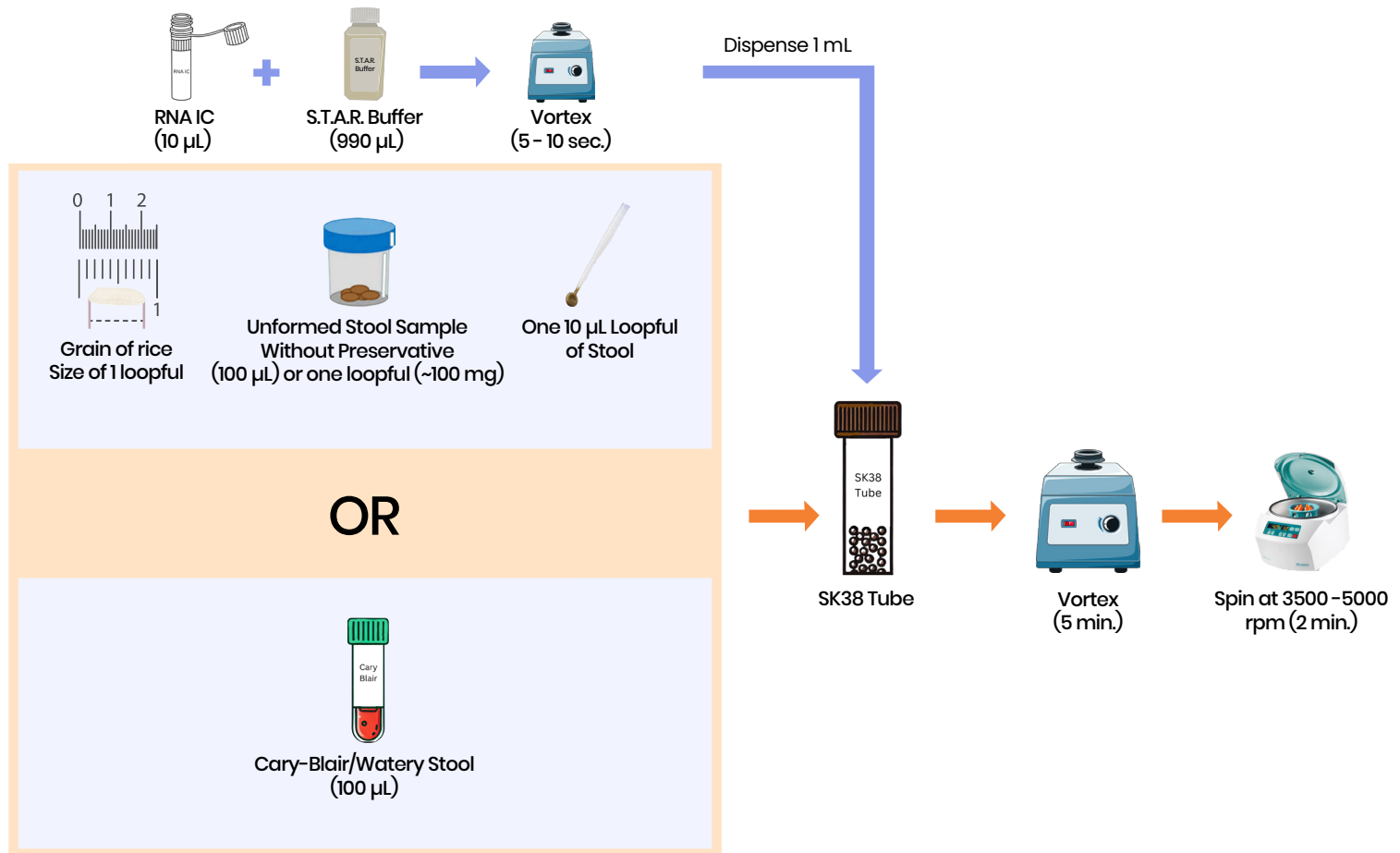
## Tips for Successful Sample Prep and Extraction With MagNA Pure 96

### Stool Amount:

Mix RNA IC and S.T.A.R. Buffer at a 1/100 ratio (v/v) to obtain 1 mL of solution for each specimen plus a negative control. Vortex for 5-10 seconds, then dispense into the SK-38 tubes. **Add 100 µL Cary-Blair** or watery stool or one loopful (~100 mg) of formed stool to the SK38 tubes. Use 10 µL-Loop to pick up a loop of formed stool to add to SK38 tube. For the negative control, add 100 µL clean media (i.e. S.T.A.R. Buffer) or well characterized negative sample. Do not add more stool than instructed. Doing so may lead to “invalid results”.

### Note:

A valid negative control is **required** for each plate/kit lot to obtain results.



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## Extraction:

Transfer 200 µL of lysate from the SK38 tube into a MagNA Pure 96 processing cartridge.

### MagNA Pure 96

#### Perform Protocol:

Pathogen Universal 200 4.0 or higher for MagNA Pure Kit: DNA/Viral NA SV 2.0

**Volume:** 200 µL

**Eluate:** 50 µL

#### Note

- Be careful to pipette directly to the bottom without producing bubbles.
- Liquid on the side of the well and bubbles will lead to incorrect volume sensing and the extraction will be aborted.

## Nucleic Acid Storage Conditions:

Optional: Transfer sample extracts from the cartridge into PCR grade container.

### 2–8°C refrigerator



If testing **within** 24 hours.

### – 80°C or below



If testing **cannot** be completed within 24 hours of extraction.

#### Note

- Store extracted nucleic acids at –80°C or below for up to 90 days.
- Store leftover pretreated samples (in SK38 tubes) at –80°C or below for up to 90 days.

## Repeat/Reflex Extraction:



MagNA Pure 96

- Transfer **50 µL** from the SK38 tube and 150 µL S.T.A.R. buffer into a MagNA Pure 96 processing cartridge.
- **Perform Protocol:** Pathogen Universal 200 4.0 or higher for MagNA Pure Kit: DNA/Viral NA SV 2.0. Volume: 200 µL, Eluate: 50 µL.

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